

Methods for neural circuit inference from population calcium imaging data

Liam Paninski

Department of Statistics and Center for Theoretical Neuroscience
Columbia University

<http://www.stat.columbia.edu/~liam>

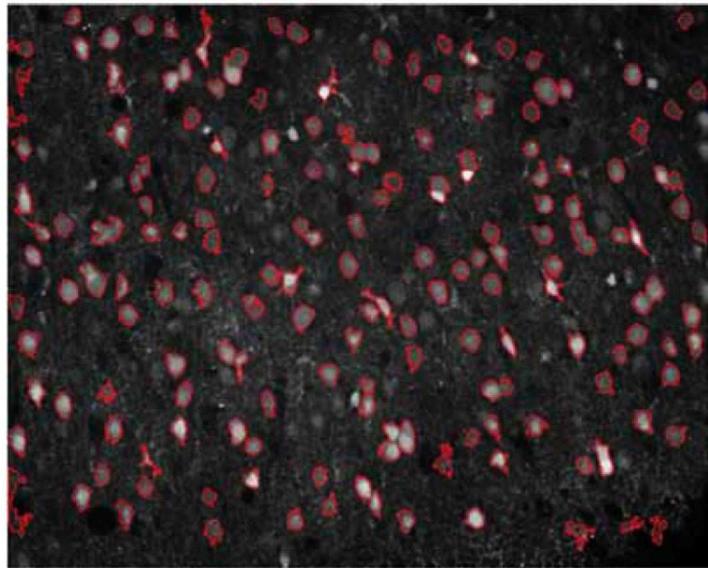
liam@stat.columbia.edu

June 6, 2010

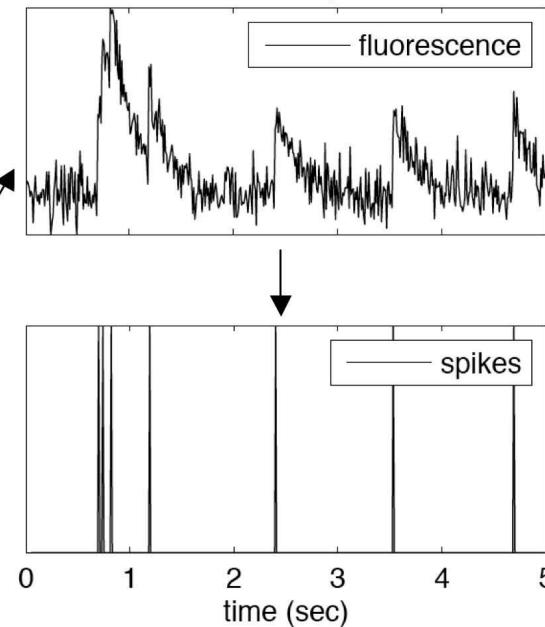
co-PI: R. Yuste (Columbia)

— with S. Koyama (CMU), **J. Vogelstein** (JHU), **Y. Ahmadian**, B. Babadi, **T. Machado**, **Y. Mishchenko**, A. Packer, K. Rahnama Rad, T. Sippy (Columbia).

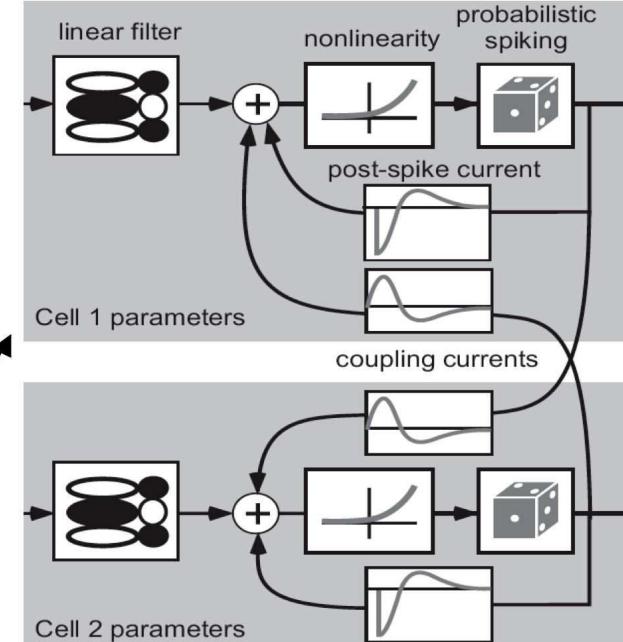
Record large-scale calcium movie



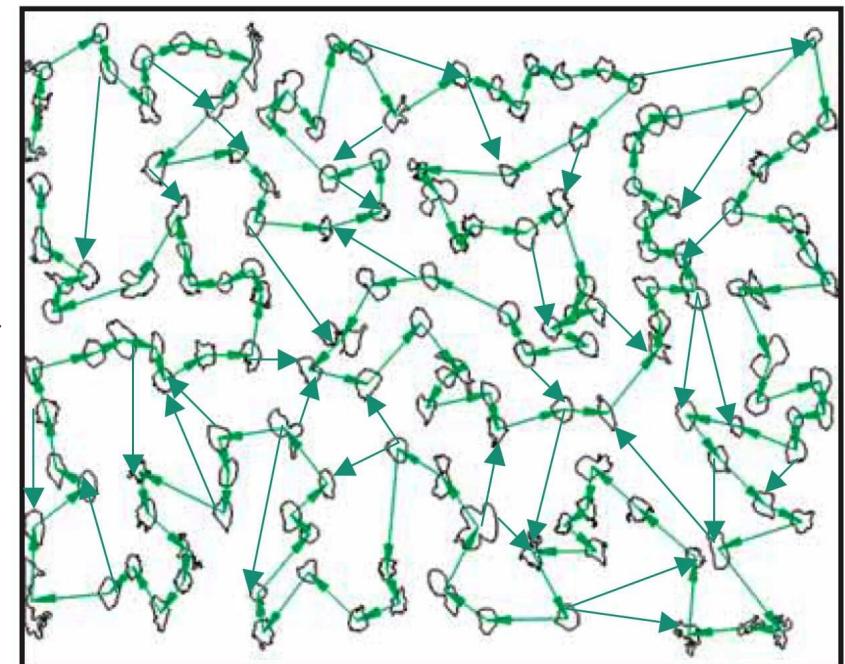
Aim 1: Extract spike times



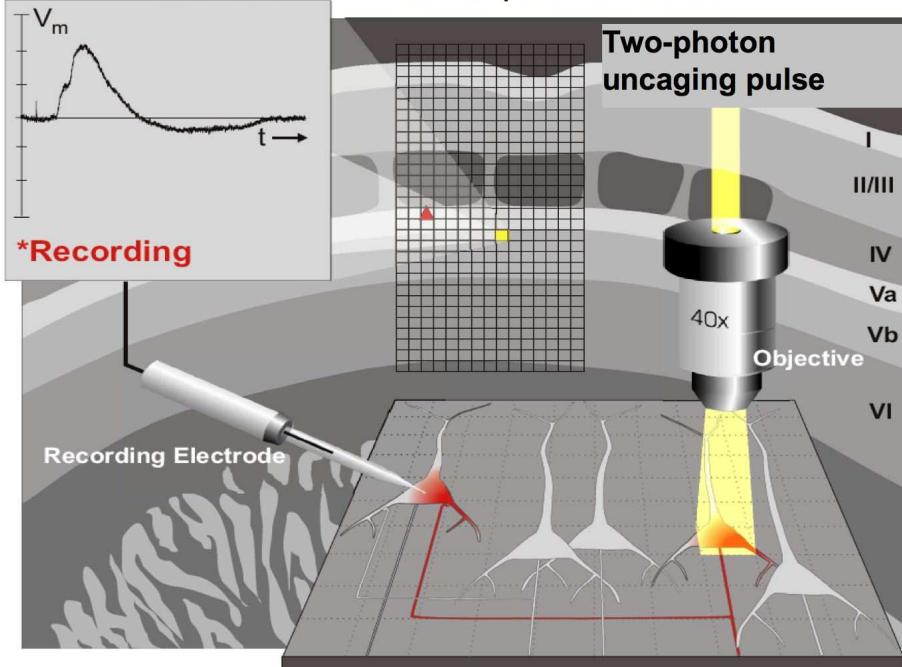
Aim 2: Estimate network model



Inferred network model

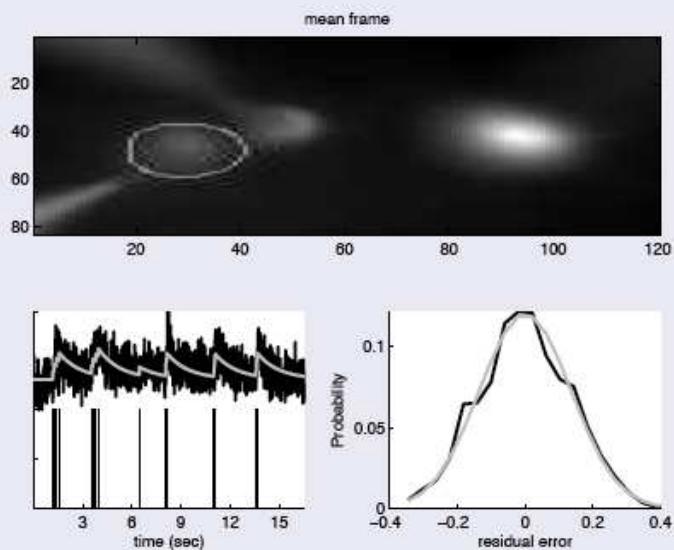


Aim 3: Check results via photostimulation

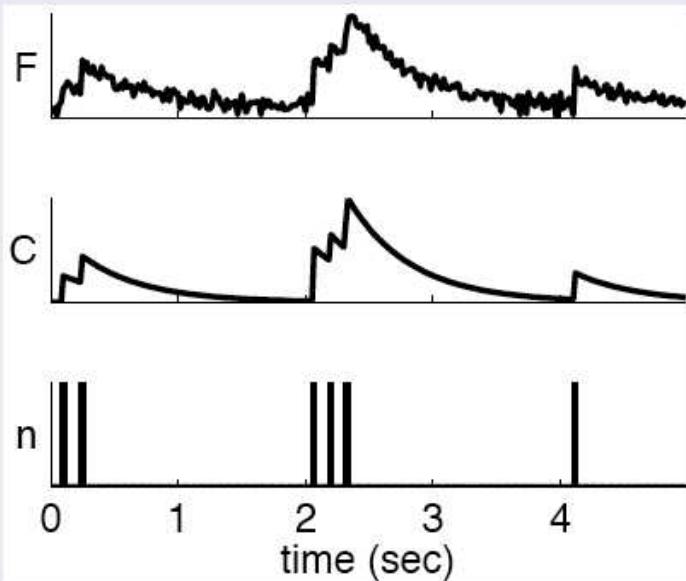


Aim 1: Model-based estimation of spike rates

data



schematic



equations

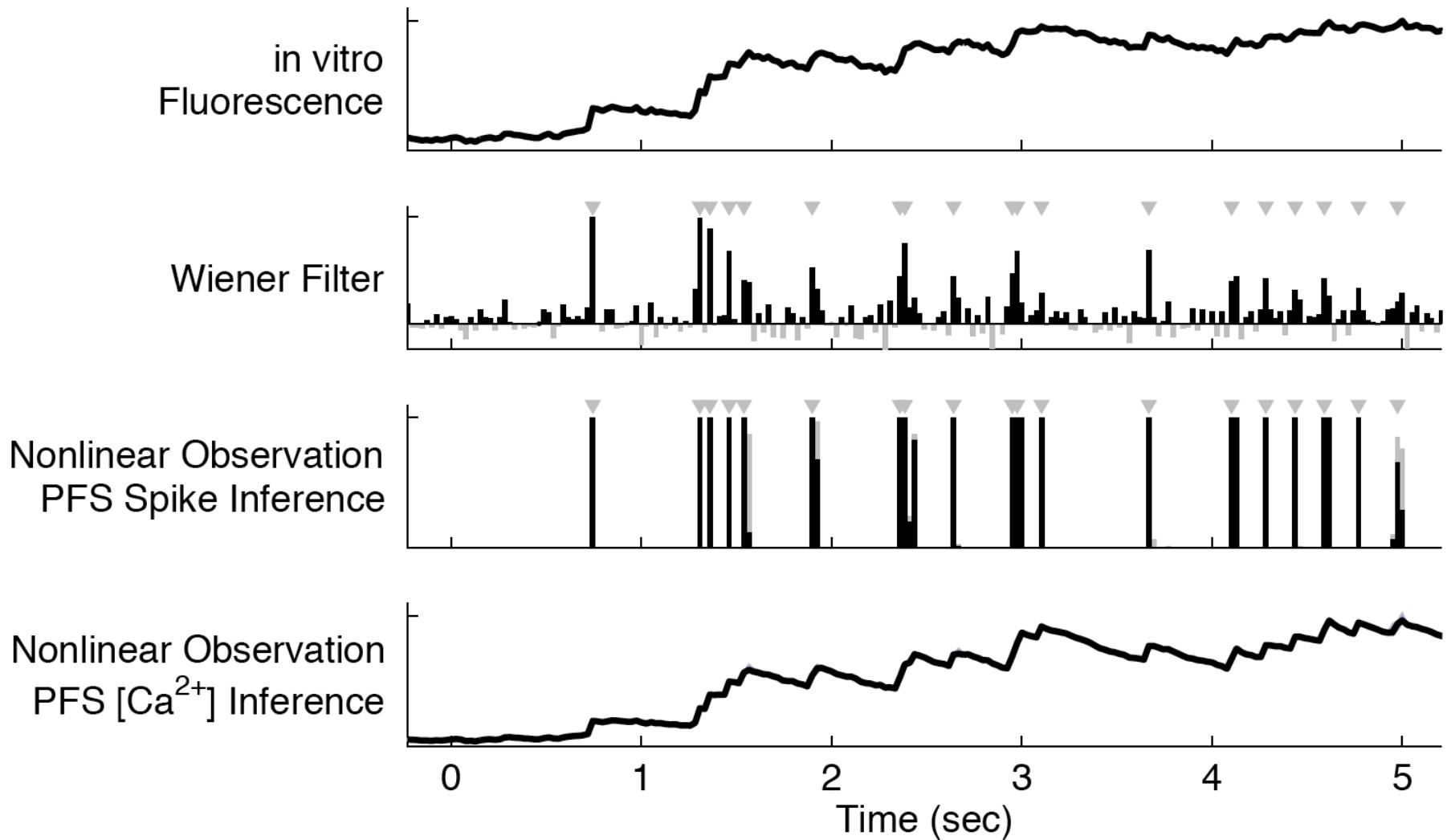
$$F_t = \alpha C_t + \beta + \sigma \varepsilon_t, \quad \varepsilon_t \stackrel{iid}{\sim} \mathcal{N}(0, 1)$$

$$C_t = -(1 - \Delta/\tau) C_{t-1} + n_t$$

$$n_t \sim \text{poisson}(\lambda \Delta)$$

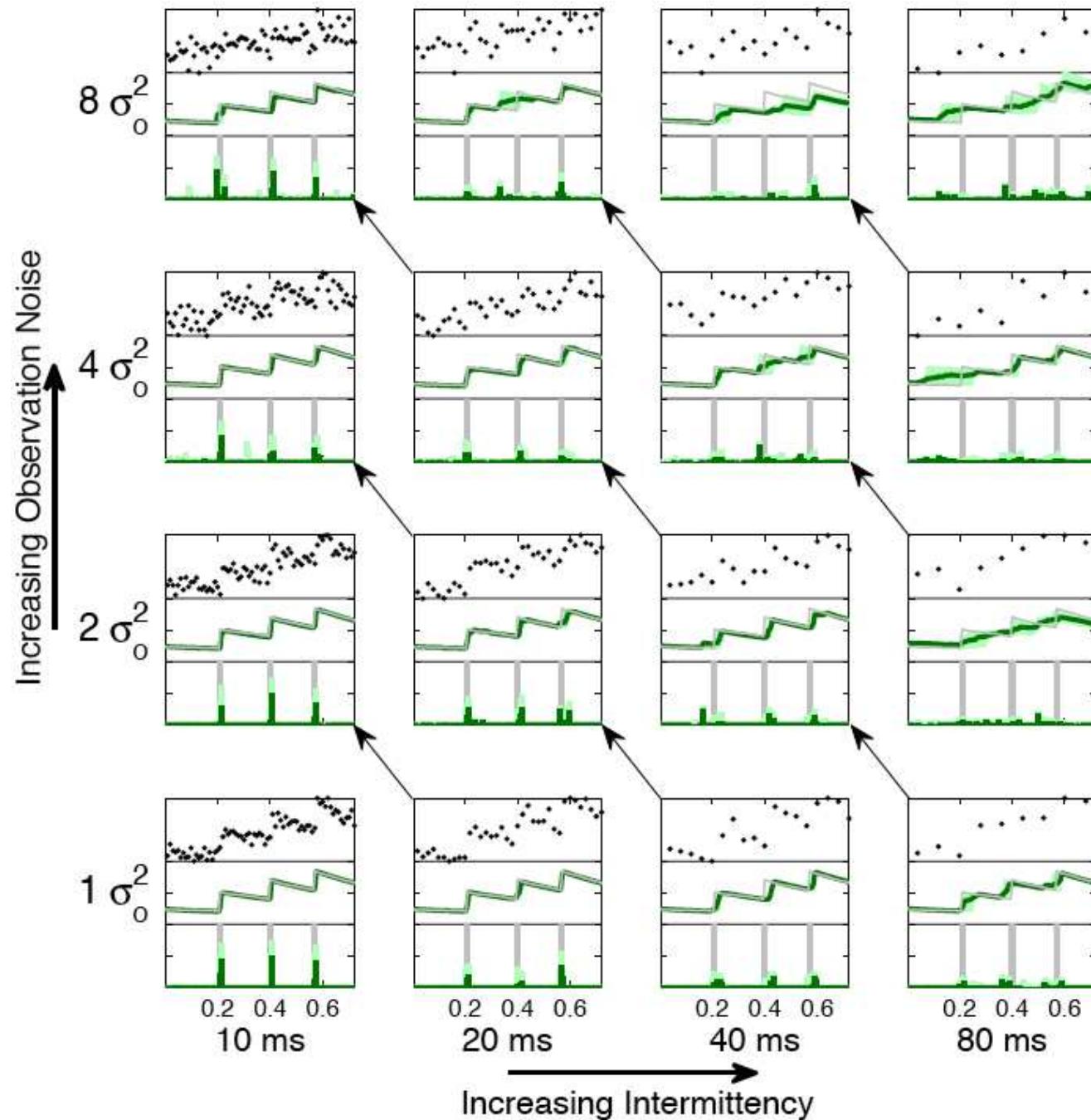
Note: each component here can be generalized easily.

Particle filter can extract spikes from saturated recordings



Optimal nonlinear filter given model; runs in linear time (like optimal linear filter).
Parameters inferred via expectation-maximization: no need for intracellular calibration experiments (Vogelstein et al., 2009).

Experimental design: speed vs. SNR tradeoff



Another look: fast maximum a posteriori (MAP) optimization

In standard linear filtering setting, forward-backward recursions also compute MAP (because $E(n|F)$ and $\hat{n} = \arg \max_n p(n|F)$ coincide if $p(n|F)$ is Gaussian).

More generally, write out the posterior:

$$\begin{aligned}\log p(C|F) &= \log p(C) + \log p(F|C) + \text{const.} \\ &= \sum_t \log p(C_{t+1}|C_t) + \sum_t \log p(F_t|C_t) + \text{const.}\end{aligned}$$

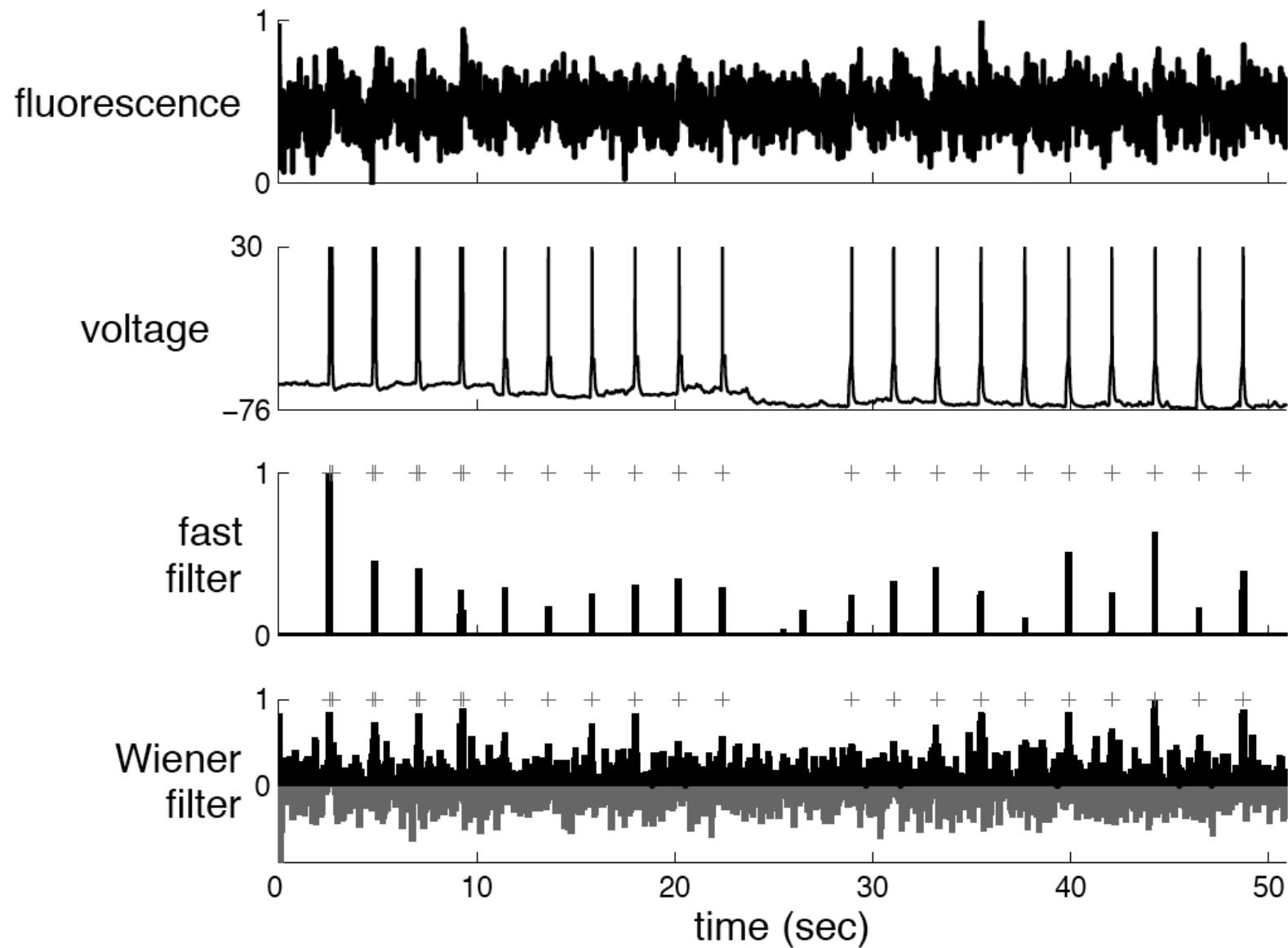
Three basic observations:

- If $\log p(C_{t+1}|C_t)$ and $\log p(F_t|C_t)$ are concave, then so is $\log p(C|F)$.
- Hessian H of $\log p(C|F)$ is tridiagonal: $\log p(F_t|C_t)$ contributes a diag term, and $\log p(C_{t+1}|C_t)$ contributes a tridiag term (Paninski et al., 2010).
- C is a linear function of n .

Newton's method: iteratively solve $HC_{dir} = \nabla$. Tridiagonal solver requires $O(T)$ time. Can include nonneg constraint $n_t \geq 0$ (Koyama and Paninski, 2009).

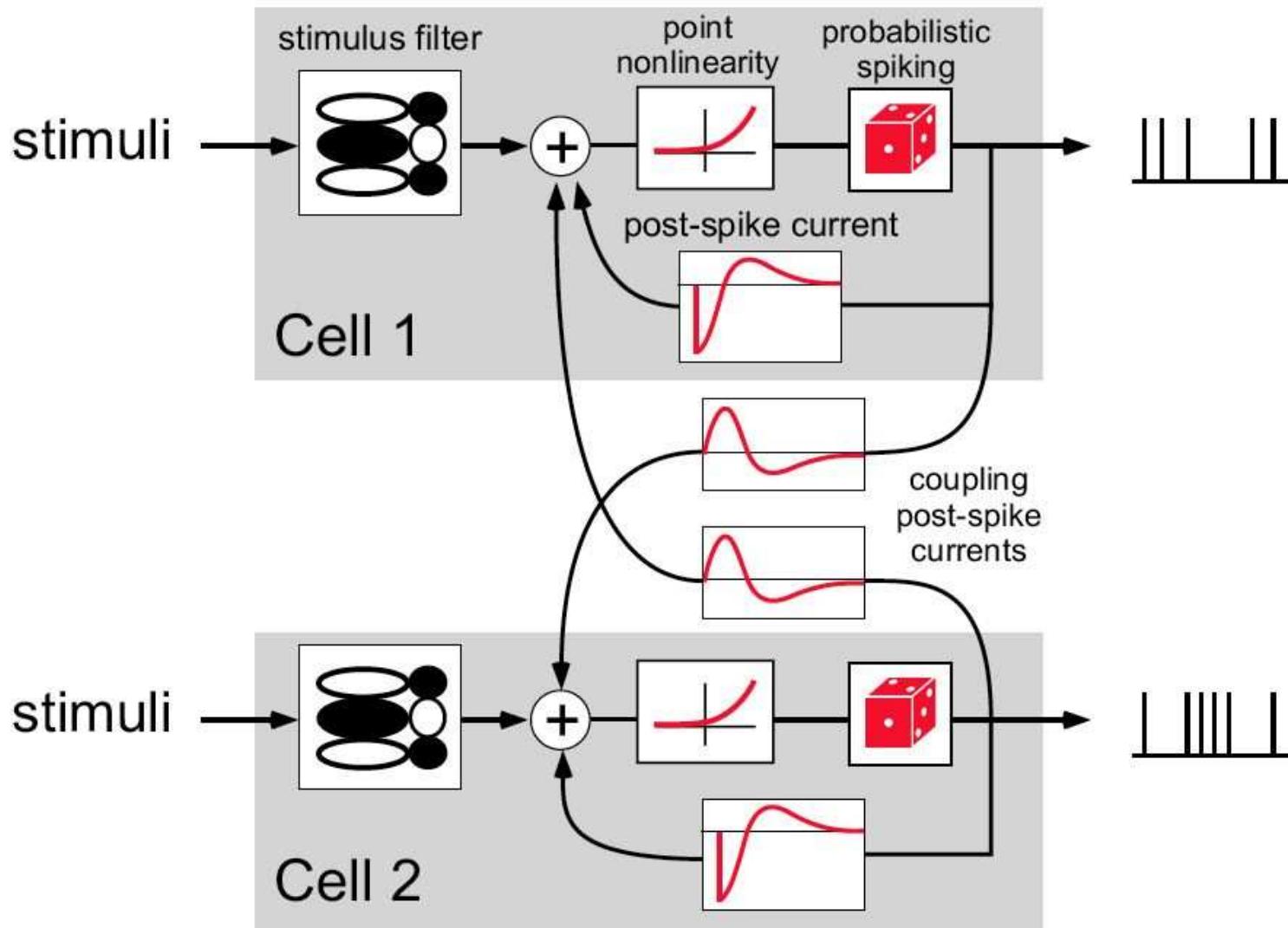
— **Two orders of magnitude faster** than particle filter: can process data from ≈ 100 neurons in real time on a laptop (Vogelstein et al., 2010).

Example: nonnegative MAP filtering



— nonnegative deconvolution is a recurring problem in signal processing
(Vogelstein et al., 2010).

Aim 2: estimating network connectivity



Given the spike times in the network, L₁-penalized concave loglikelihood optimization is easy (Paninski, 2004; Pillow et al., 2008). Fast, efficient methods from generalized linear model, compressed sensing literature.

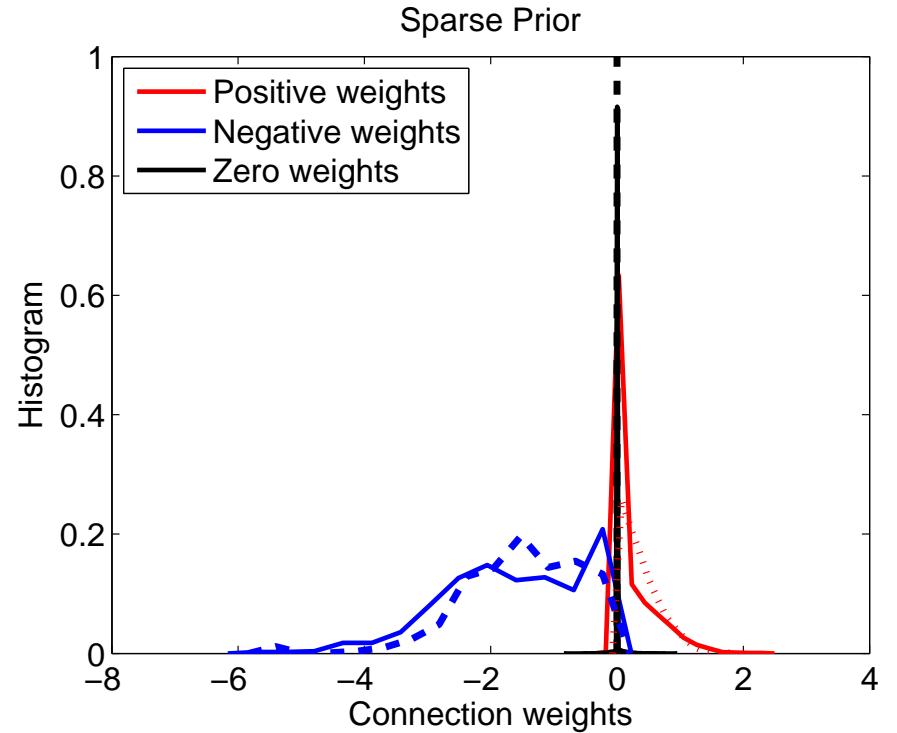
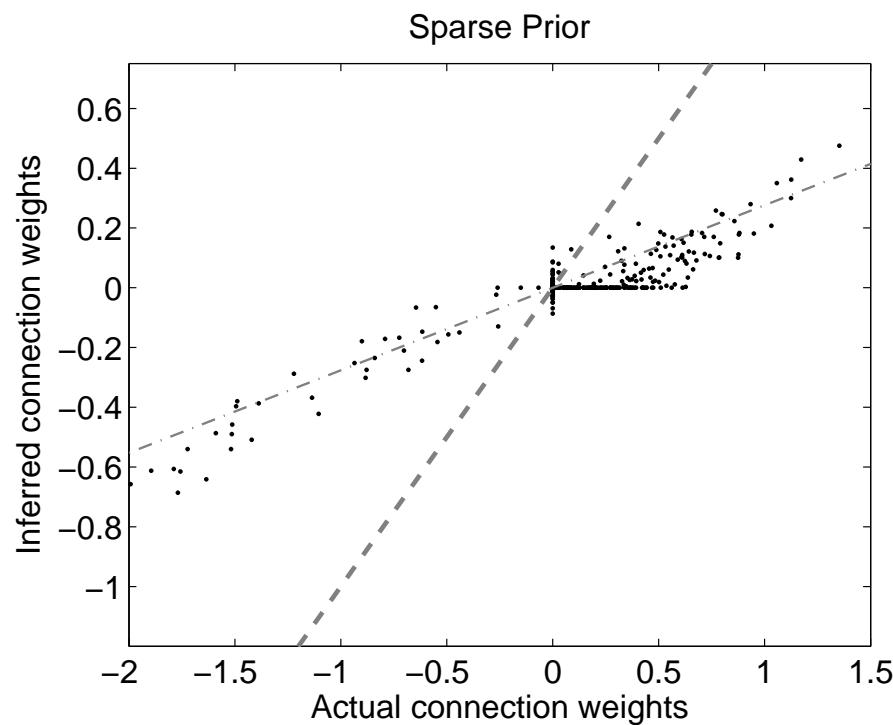
Monte Carlo EM approach

...But we only have noisy calcium observations F ; true spike times are hidden variables. Thus an EM approach is once again natural.

- E step: sample spike train responses n from $p(n|F, \theta)$
- M step: given sampled spike trains, perform L_1 -penalized likelihood optimization to update parameters θ .

E step is hard part here. Use the fact that neurons interact fairly weakly; thus we need to sample from a collection of weakly-interacting Markov chains, via Metropolis-within-blockwise-Gibbs forward-backward methods (Mishchenko et al., 2010).

Simulated circuit inference



— conductance-based integrate-and-fire networks with biologically plausible connectivity matrices, imaging speed, SNR (Mishchenko et al., 2009).

Good news: MAP connections are inferred with the correct sign, in just a couple minutes of compute time. Current work focusing on improved sampling methods, to better quantify uncertainty (exploiting hybrid forward-backward blockwise-Gibbs approach) (Mishchenko and Paninski, 2010).

Aim 3: Optimal control of spike timing

To test our results, we want to perturb the network at will.
How can we make a neuron fire exactly when we want it to?

Assume bounded inputs; otherwise problem is trivial.

Start with a simple model:

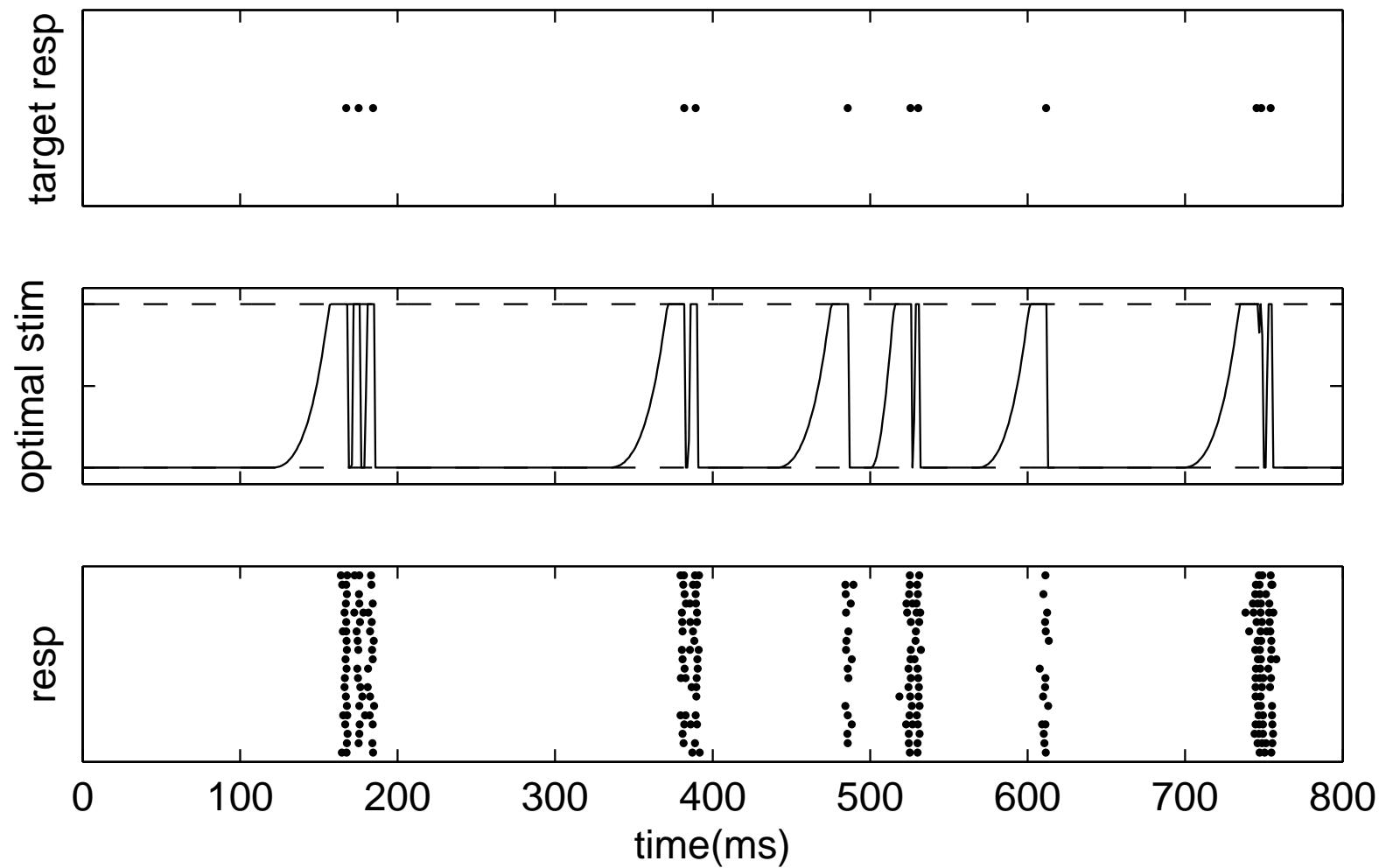
$$\begin{aligned}\lambda_t &= f(V_t + h_t) \\ V_{t+dt} &= V_t + dt(-gV_t + aI_t) + \sqrt{dt}\sigma\epsilon_t, \quad \epsilon_t \sim \mathcal{N}(0, 1).\end{aligned}$$

Now we can just optimize the likelihood of the desired spike train, as a function of the input I_t , with I_t bounded.

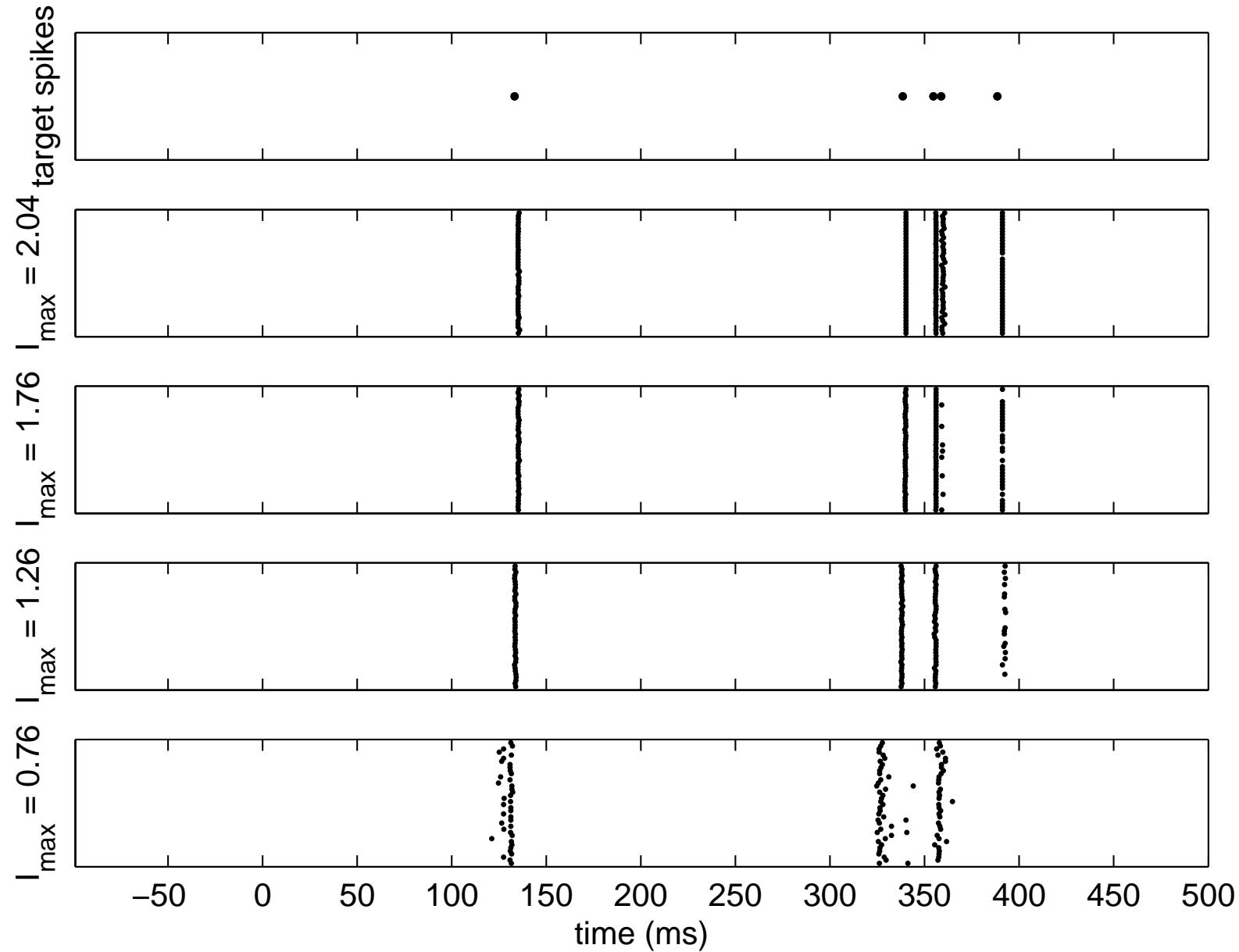
Concave objective function over convex set of possible inputs I_t
+ Hessian is tridiagonal $\implies O(T)$ optimization.

— again, can be done in real time (Ahmadian et al., 2010).

Simulated electrical control of spike timing



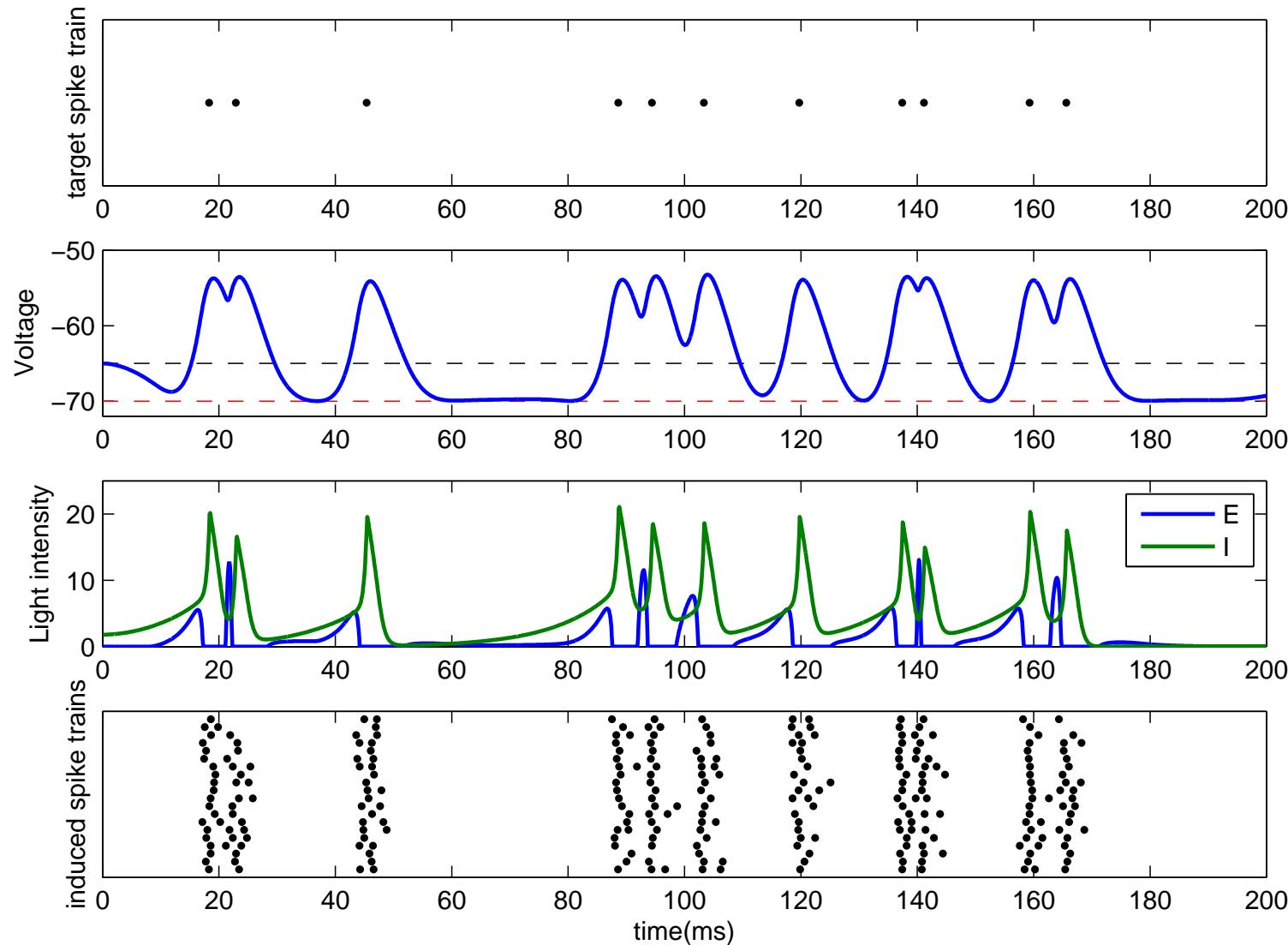
Example: intracellular control of spike timing



(Ahmadian et al., 2010)

Optical conductance-based control of spiking

$$\begin{aligned}
 V_{t+dt} &= V_t + dt \left(-gV_t + g_t^i(V^i - V_t) + g_t^e(V^e - V_t) \right) + \sqrt{dt}\sigma\epsilon_t, \quad \epsilon_t \sim \mathcal{N}(0, 1) \\
 g_{t+dt}^i &= g_t^i + dt \left(-\frac{g_t^i}{\tau_i} + a_{ii}L_t^i + a_{ie}L_t^e \right); \quad g_{t+dt}^e = g_t^e + dt \left(-\frac{g_t^e}{\tau_e} + a_{ee}L_t^e + a_{ei}L_t^i \right)
 \end{aligned}$$



Next steps

- Optimize experimental parameters: imaging speed, number of neurons, scanning in 3D. Spatial light modulation microscopy a key tool (Nikolenko et al., 2008).
- Stimulate network electrically and optically to increase number of observed distinct spiking patterns: Shepard 256 × 256 electrode array, channelrhodopsin, new uncaging compounds (Rial Verde et al., 2008)
- Incorporate hidden neuron effects in analysis (Vidne et al., 2009)
- Check results with genetically-labeled inhibitory cells, dual patching, optical stimulation of putative presynaptic neurons (Nikolenko et al., 2007)
 - Postdocs wanted!

References

- Ahmadian, Y., Packer, A., Yuste, R., and Paninski, L. (2010). Fast optimal control of spike trains. *Under review.*
- Koyama, S. and Paninski, L. (2009). Efficient computation of the map path and parameter estimation in integrate-and-fire and more general state-space models. *Journal of Computational Neuroscience*, In press.
- Mishchenko, Y. and Paninski, L. (2010). Efficient methods for sampling spike trains in networks of coupled neurons. *In preparation.*
- Mishchenko, Y., Vogelstein, J., and Paninski, L. (2010). A Bayesian approach for inferring neuronal connectivity from calcium fluorescent imaging data. *Annals of Applied Statistics*, In press.
- Nikolenko, V., Poskanzer, K., and Yuste, R. (2007). Two-photon photostimulation and imaging of neural circuits. *Nature Methods*, 4:943–950.
- Nikolenko, V., Watson, B., Araya, R., Woodruff, A., Peterka, D., and Yuste, R. (2008). SLM microscopy: Scanless two-photon imaging and photostimulation using spatial light modulators. *Frontiers in Neural Circuits*, 2:5.
- Paninski, L. (2004). Maximum likelihood estimation of cascade point-process neural encoding models. *Network: Computation in Neural Systems*, 15:243–262.
- Paninski, L., Ahmadian, Y., Ferreira, D., Koyama, S., Rahnama, K., Vidne, M., Vogelstein, J., and Wu, W. (2010). A new look at state-space models for neural data. *Journal of Computational Neuroscience*, In press.
- Pillow, J., Shlens, J., Paninski, L., Sher, A., Litke, A., Chichilnisky, E., and Simoncelli, E. (2008). Spatiotemporal correlations and visual signaling in a complete neuronal population. *Nature*, 454:995–999.
- Rial Verde, E., Zayat, L., Etchenique, R., and Yuste, R. (2008). Photorelease of GABA with visible light using an inorganic caging group. *Frontiers in Neural Circuits*, 2:2.
- Vidne, M., Kulkarni, J., Ahmadian, Y., Pillow, J., Shlens, J., Chichilnisky, E., Simoncelli, E., and Paninski, L. (2009). Inferring functional connectivity in an ensemble of retinal ganglion cells sharing a common input. *COSYNE*.
- Vogelstein, J., Packer, A., Machado, T., Sippy, T., Babadi, B., Yuste, R., and Paninski, L. (2010). Fast non-negative deconvolution for spike train inference from population calcium imaging. *J. Neurophys.*, In press.
- Vogelstein, J., Watson, B., Packer, A., Jedynak, B., Yuste, R., and Paninski, L. (2009). Model-based optimal inference of spike times and calcium dynamics given noisy and intermittent calcium-fluorescence imaging. *Biophysical Journal*, 97:636–655.